

# Plant-Environment Interaction

Responses and Approaches to Mitigate Stress

Edited by  
Mohamed Mahgoub Azooz • Parvaiz Ahmad



**WILEY** Blackwell

## **Water Stress and Crop Plants**

# **Water Stress and Crop Plants**

A Sustainable Approach,  
Volume 1

EDITED BY

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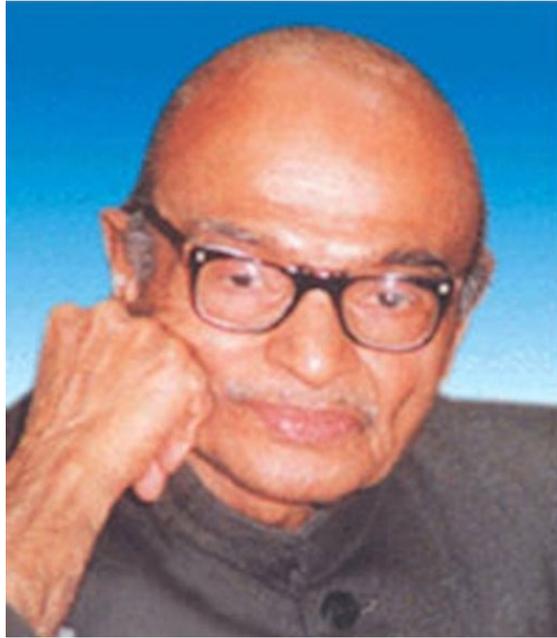
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Dedicated  
to



**Hakim Abdul Hameed**  
(1908–1999)  
Founder of Jamia Hamdard  
(Hamdard University)  
New Delhi, India

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# Foreword

Humans started their community life nearly 10,000 years back by beginning to gather and cultivate plants and domesticate animals. In this way the foundations for agriculture were laid as an important part of life. A great development has taken place since then, but still a large population is suffering from hunger in different countries. Land degradation is leading to tremendous soil losses and different types of stresses are posing great threat to the soil productivity, which in turn is affecting plant growth and development ending up with decreases in the crop yields.

On the other hand, demographic developments are posing another threat and attempts are to be made to combat this grave situation in order to feed the hungry. Plant scientists are trying hard to develop plants with higher yields and those which can be grown on marginal lands. They are working hard to develop techniques with latest technologies to understand the molecular, physiological, and biochemical pathways in order to meet the global agricultural needs by overcoming the stresses affecting the yield.

Water is the most critical resource for a sustainable agricultural development in the world. It is a must for the agriculture as an important part of our environment. The problems arising from under and overirrigation emphasize the fact that humans cannot continue with the current use and throw away policy with their natural resources; in particular, regarding water. The area of irrigated lands is reaching a level of nearly 500 million ha and approximately 20% of these irrigated lands provide only 50% of the global food supply. Expectations are that the need for irrigation water will increase far more by 2025. Water scarcity will cause stress problems in plants. In view of this we have to look for the possibilities to overcome water shortages in the agriculture so as to increase the water use efficiency, use marginal lands, marginal waters, and techniques to overcome stress problems in plants to feed hungry mouths.

This volume is therefore a compilation of different perspectives from around the globe that directly or

indirectly lead us to understand the mechanism of plant stress tolerance and mitigation of these dangerous stresses through sustainable methods.

Chapter 1 deals with the drought stress and photosynthesis in plants. Here, the authors give details regarding the effect of drought on photosynthesis in plants, stomatal and non-stomatal limitation of photosynthesis during drought stress, resistance of plants to drought stress, and effect of drought stress on leading plants.

Chapter 2 discusses the role of crassulacean acid metabolism induction in plants as an adaptation to water deficit; physiological and metabolic aspects of CAM induction by drought, CAM induction and fitness under water deficit; capability of CAM to improve water-use efficiency, and productivity is also explained clearly.

In Chapter 3 authors enlighten the effect of drought stress on the functioning of stomata, and hormonal, nutritional, as well as genetic aspects under drought stress.

Chapter 4 discusses the case study under the heading of recurrent droughts with details about keys for sustainable water management from case studies of tree fruit orchards in central Chile.

In Chapter 5, global explicit profiling of water deficit-induced diminutions in agricultural crop sustainability is given as a key emerging trend and challenge; defensive mechanisms adopted by crops at whole plant level under specific drought scenarios: perception, sensing, and acclimation is also explained.

The information on sustainable agricultural practices for water quality protection are discussed at length in Chapter 6.

In Chapter 7, salinity and drought stress topics are evaluated including information on the similarities and differences in oxidative responses and cellular redox regulation; similarities and differences in ROS metabolism under salinity and drought, together with water stress  $\times$  salt stress effects on plants and possible tolerance mechanisms.

The oxidative stress and plant responses to pathogens under drought conditions are discussed at length in Chapter 8.

In Chapter 9, the potential use of antioxidants, hormones, and plant extracts are reviewed with innovative approaches in taming water stress limitation in crop plants; the authors stress upon the impact of water stress on growth and development, yield, physiological processes, oxidative stress, adaptation strategies, application for osmoprotectants, and plant extracts as antioxidants.

The main topics reviewed in Chapter 10 are water stress in plants, from genes to biotechnology, identifying the genes associated with drought tolerance and engineering drought tolerance.

Chapter 11 analyzes plant aquaporins in abiotic stress tolerance under such headings as; status and prospects, functional diversity of aquaporins in plants, aquaporin gene expression studies under abiotic stresses, and genetic manipulation of aquaporin functions in transgenic plants.

Chapter 12 presents a discussion on the role of proteins in alleviating drought stress in plants, with information on functional and regulatory proteins, QTL analysis, and breeding.

The avenues for improving drought tolerance in crops by ABA regulation with molecular and physiological basis are debated in Chapter 13; whereas MYB transcription factors for enhanced drought tolerance in plants are given in Chapter 14. Here, it also explains the molecular responses to stress, transcription factors – major players in the control of gene expression and MYB transcription factors in drought stress.

Chapter 15 presents an overview dealing with the analysis of novel haplotype variations at *TaDREB-D1* and *TaCwi-D1* genes influencing drought tolerance in bread/synthetic wheat derivatives.

The TFs, master switches with multiple roles in regulatory networks for abiotic stress tolerance, transgenic plants harboring TFs versus drought stress tolerance,

microRNAs and drought stress tolerance, a fact or fiction and systems-based approach for functional genomics in plants is discussed at length in Chapter 16.

Chapters 17 and 18 deal with the role of miRNA/siRNA to enhance drought tolerance of barley and wheat and other crops; whereas Chapter 19 demonstrates sugar signaling in plants, a novel mechanism for drought stress management together with the role of sugars, osmoregulation under drought stress, sugars as signaling molecules, and exogenous application of sugars to alleviate the drought stress.

In Chapter 20, information on agriculture, socio-economic, and cultural relevance of wild relatives of crops, in particular, food legume landraces, in Northern Africa, are well documented.

I am sure that this volume will be beneficial to the students as well as staff of agricultural faculties, agricultural engineers working in the extension services, environmentalists, and also for agro-industry workers. I extend my deepest appreciations to the editor as well as the contributors for the hard labor they have put in producing this excellent volume.

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# Preface

Water stress is accepted as one of the major abiotic stresses faced on a global scale. The reasons for this could be less availability of water, which results in drought, or presence of excessive amount of water leading to waterlogging. Drought as well as waterlogging have negative impacts on plant growth and development and ultimately affect the production of crops. The primary stresses imposed here are osmotic and ionic stress, however, prolonged effects can cause secondary stress known as oxidative stress. In the latter case, the generation of reactive oxygen species is evolved, which attack the biomolecules and hamper their normal functions. Although research on impact of water stress on plants is going at high speed at global level, the effects at biochemical and molecular levels are still unclear. To understand the physiological, biochemical, and molecular mechanisms involved in environmental stress perception, transduction, and tolerance is still a challenge facing plant biologists.

Plants are equipped with different resistance mechanisms to survive under these harsh conditions. Scientists are investigating the possibilities to create water resistant crops to bring the marginal lands in to cultivation so that growing population can meet the hunger need. The current book entitled *Water Stress and Crop Plants: A Sustainable Approach* has two volumes covering all aspects of drought and flooding stress, causes and

consequences, mitigation of water stress, modern tools, and techniques to alleviate water stress and production of crop yields under water stress. The first volume includes 20 chapters enlightening the reader to different aspects with the latest knowledge and provides extensive information regarding the crop plants, their growth and development, physiological and molecular responses, together with the adaptability of crop plants to different environmental stresses.

Chapters contributed here have been published whilst keeping intact author's justifications; however, suitable editorial changes have been incorporated wherever considered necessary. We have tried our best to gather the information on different aspects of this volume, however, there is a possibility that some errors still creep in to the book for which we seek reader's indulgence and feedback. We are thankful to the authors for their valuable contributions and to John Wiley & Sons, Ltd, Chichester, particularly Gudrun Walter (Editorial Director, Natural Sciences), Audrie Tan (Project Editor), Laura Bell (Assistant Editor), and all other staff members at Wiley, who were directly or indirectly associated with us in this project for their constant help, valuable suggestions, and efforts in bringing out the timely publication of this volume.

**Parvaiz Ahmad**

## CHAPTER 11

# Lipid metabolism and oxidation in plants subjected to abiotic stresses

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### 11.1 Introduction

Plant lipids include fats, waxes, steroids, phospholipids and hydrocarbons. The free higher fatty acids and their salts (soaps) also belong to this category. Below are detailed the main classes of plant lipids.

#### 11.1.1 Membrane lipids

These are important chemical components of all cell membranes, where they are represented mainly by amphipathic phospholipids (the more abundant) and sterols (particularly stigmasterol), spontaneously forming bilayers in an aqueous environment (López *et al.*, 2011). The exception is the thylakoid membranes of chloroplasts, which contain primarily galactolipids, that are also amphipathic and mostly stable in a bilayer configuration (Robinson & Mant, 2005; Tetlow *et al.*, 2010). The amphipathic nature of membranes permits the formation of membranous sheets that self-anneal their edges into a sealed compartment. The inner and outer surfaces of a membrane, both plasma and organelle membranes, differ considerably in chemical composition (Evert, 2006).

#### 11.1.2 Carotenoids

These are yellow to red-coloured non-polar lipids with a terpenoid structure containing eight isoprene units arranged in a symmetrical linear pattern. Their conjugated double bonds are responsible for the typical visible light absorption of this class of compounds. Carotenoids are responsible for protecting the chlorophylls from photo-dynamic destruction (Cuttriss & Pogson, 2004). They

exist in many flowers and fruits where they are valuable in terms of providing signals to other species on the availability of food sources, and in this way ensure pollination and the spread of seeds.

The two major classes of carotenoids are the carotenes and their oxygenated derivatives, the xanthophylls. The most abundant xanthophylls, lutein and violaxanthin, are key components of the light-harvesting complex (LHC) of leaves (Foyer & Shigeoka, 2011; Singh & Tuteja, 2010).

#### 11.1.3 Oil bodies

Oil bodies are spherical structures, also called spherosomes or oleosomes, particularly concentrated in fruits and seeds, arising by the accumulation of triacylglycerol molecules. Seeds contain high amounts of lipids, together with proteins and carbohydrates, accumulated as a nutrient source for seedling growth. Approximately 45% of the weight of sunflower, peanut, flax and sesame seeds is composed of oil (Bradford & Nonogaki, 2007). Lipids of seed cells have been suggested to be sensitive to temperature, thus functioning as a 'temperature probe' able to detect the signals for stopping dormancy. Aleurone grains within endosperm store abundant reserves of lipids that are broken down to sugar by beta-oxidation and the glyoxylate cycle to fuel hypocotyl elongation in the dark (Golovina & Hoekstra, 2003). The outer layers of seed integument are covered and/or impregnated with more or less solid lipids which are primarily responsible for seed waterproofing properties. When the seeds of various species of Sapindaceae are injured by pathogens, specific lipids (cyanogenic

lipids) are rapidly cleaved by an esterase, liberating HCN as a defence response (Selmar, 2010). The essential oils, present mainly in the leaves of some species (e.g. Lamiaceae), are volatile oils that contribute to the essence and odour of plants. They are made by special cells in the form of oil bodies and then excreted into intercellular cavities. Oil bodies are also common storage products and cryoprotectants of dormant cambial cells (Evert, 2006).

#### 11.1.4 Waxes

Waxes are long-chain (20–25 C) lipid compounds (mixtures of hydrocarbon molecules, predominantly paraffins and related alcohols, ketones, acids and esters), solid at normal temperatures, constituting part of the protective coating (cuticle) on the epidermis of the aerial parts of the primary plant body and on the inner surface of the primary wall of cork cells in woody roots and stems. They constitute a major barrier to water loss from the surface of the plant by reducing the wetability of leaves. They also reduce the ability of fungal spores to germinate and of bacteria to grow, thereby reducing the ability of these agents to cause diseases (Zentgraf, 2007). Cuticle components are made in the epidermal cells and are extruded through the outer walls to the exterior, where they take up their final form. In the nectaries of many flowers, the nectar is released by rupture of the wall and cuticle of each epidermal cell (Evert, 2006). The nectar has high sugar content, mainly sucrose, and also contains lipids and proteins.

The smooth endoplasmic reticulum (SER) is involved in the synthesis of different types of lipidic compound (Evert, 2006). The role of the SER labyrinths is not fully understood but it is present in a variety of plant glands, secreting fats, oils and fragrant essential oils, and is also abundant in epidermal cells that are making lipid molecules to be deposited in the external cell wall waxes, cutin and suberin (Robinson & Mant, 2005). In general, the SER has been found to process the enzymes necessary for making complex lipids, given the ingredients of fatty acids (made in plastids) and lipid head groups (made by cytoplasmic enzymes). On the basis of the model best supported by experimental evidence, these raw materials come together in the cytoplasmic face of the SER membrane to produce a variety of products. Depending on their nature, they may accumulate until they form lipid droplets, initially still in the membrane but then liberated into the

cytoplasm for storage or transport around the cell. Special enzymes are able to ‘flip’ lipids from the cytoplasmic face of the ER membrane (where they are made) to the luminal face, correcting the imbalance that arises from asymmetrical synthesis. In this way, the ER membrane grows in surface area. Expanses of membrane may then be mobilized to other systems in the form of vesicles – especially to the Golgi apparatus and from there to the plasma membrane or other cell parts (Tetlow *et al.*, 2010).

Whereas most membranes are composed primarily of phospholipids, galactolipids account for over 80% of thylakoid membranes but they are also present in outer and inner chloroplast envelopes. Galactolipids are synthesized at the inner envelope of the chloroplast and these lipids are furthermore highly unsaturated. It is concluded that an intraorganellar lipid transport system must exist that transfers lipids from their site of synthesis to the thylakoids. Thylakoid membranes are thought to be derived from invaginations of the inner membrane, as maturing chloroplasts sometimes exhibit a continuum between the inner membrane and internal membrane structures, although this continuum is not present in mature chloroplasts. It has been suggested that vesicle trafficking from the inner membrane to the thylakoids allows maintenance and regeneration of these structures in the mature chloroplast (Robinson & Mant, 2005). Furthermore, an ATP-dependent factor involved in vesicle fusion within pepper chromoplasts has been isolated and the gene cloned.

Such a ‘budding’ mechanism of thylakoid biogenesis would explain how other hydrophobic membrane components (e.g. carotenoids and galactolipids) are synthesized on the chloroplast envelope, and are able to reach the thylakoid membranes themselves. The interaction between specific proteins and galactolipids could be important for distinguishing the chloroplast from other potential target membranes inside the cell. Leucoplasts are colourless plastids that are distinct from proplastids because they have lost their progenitor function. Within this group are elaioplasts/oleoplasts, that are the sites of lipid synthesis (Tetlow *et al.*, 2010).

The biosynthetic pathway of carotenoids involves a series of desaturations, cyclizations, hydroxylations and epoxidations, commencing with the formation of phytoene and typically terminating in lutein and neoxanthin accumulation (Cuttriss & Pogson, 2004). In brief, phytoene is formed by the condensation of geranylgeranyl diphosphate by phytoene synthase.

Phytoene is subjected to four desaturation reactions by phytoene desaturase and zeta-carotene desaturase to produce tetra-cis-lycopene, which is isomerized by the carotenoid isomerase to produce all-trans-lycopene. Lycopene is cyclized twice to produce beta-carotene or once to produce alpha-carotene. The two carotenes are hydroxylated by the beta- and epsilon-hydroxylases to produce zeaxanthin and lutein, respectively. Zeaxanthin is epoxidated by zeaxanthin epoxidase to form violaxanthin, which is further modified by neoxanthin synthase to produce neoxanthin.

Carotenoids and their biosynthetic enzymes are placed in the plastids, although carotenoid biosynthetic genes are within the nuclear genome. The pathway is at least in part regulated via changes in transcription. As a consequence, the transcriptional regulation of carotenoids and also chloroplast-nuclear signalling are probably induced by various environmental stimuli, oxidative stress, redox balance and metabolite feedback regulation (Foyer & Shigeoka, 2011).

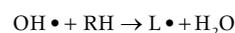
## 11.2 Lipid vulnerability to reactive oxygen species and mechanism of lipid oxidation

Lipid peroxidation starts when activated oxygen species react with the double bonds present in lipid hydrocarbon chains. On the basis of the lipids involved, type of oxidant compounds and oxidation severity, different products of lipid peroxidation are produced, such as compounds containing hydroperoxyls, hydroxyls, ketones, aldehydes, carboxylic acids and trans double bonds (Borchman & Sinha, 2002; Hameed *et al.*, 2013; Sharma *et al.*, 2012).

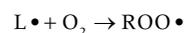
Lipid peroxidation in plants is mainly due to ROS activity. According to Browne & Armstrong (2002), the primary target on lipids of ROS is the 1,4-pentadiene structure of a polyunsaturated fatty acid (PUFA), either free or esterified to cholesterol or glycerol. The process starts (initiation) when a ROS removes a methylene hydrogen from PUFA. In this reaction, the ROS is quenched and a PUFA-centred alkoxyl radical (L•) is formed, with a consequent spontaneous rearrangement of its double bonds and the formation of a conjugated diene. Reaction of L• with oxygen in its molecular form produces a PUFA-centred peroxy radical (LOO•). The reaction continues (propagation) when either L• or

LOO• acts as initiating ROS, so attacking a neighbouring PUFA of the lipidic bilayer structure of a membrane or within a lipoprotein. Thus, a new L•, which can further propagate the reaction and form a lipid hydroperoxide (LHP), is produced. The reaction ends (termination) when an antioxidant molecule able to absorb the intermediate free radicals, or free radical scavengers, interrupts this chain reaction.

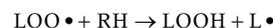
The mechanism described above can be summarized in the following phases: activation, distribution and cleavage. In brief, the activation of an unsaturated fatty acid by one radical causes the cleavage of one H<sup>•</sup> from the methyl vinyl group present in the fatty acid:



The resonance structure of this reaction reacts with triplet oxygen, a biradical with two unpaired electrons, producing a peroxide radical:

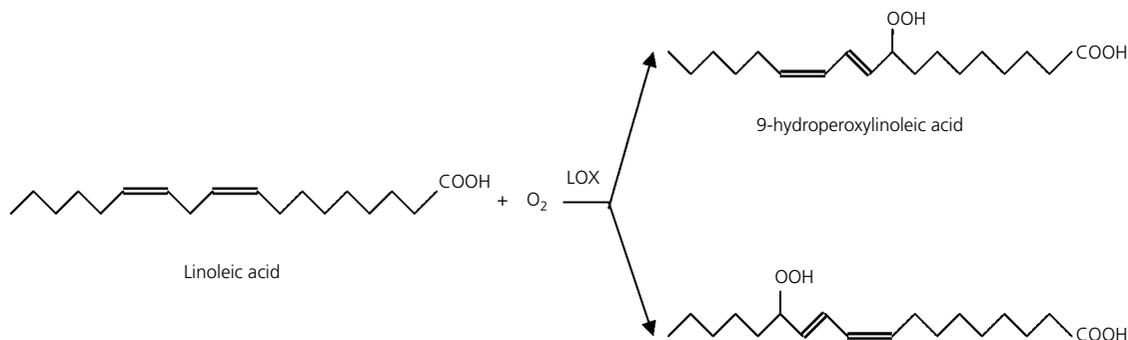


This latter reacts with a hydrogen atom from a second fatty acid, with the formation of a lipid hydroxide; the free carbon can participate in the secondary assimilation of hydrogen:



The high reactivity of hydroxide radicals in any given lipid system is due to their ability to initiate a chain reaction at very low levels (Hinojosa *et al.*, 2010).

Little is known about the ability of plants to repair the effects of stress-induced oxidative damage in cell membranes. Particularly, lipid peroxidation occurring during abiotic stresses generates changes in fatty acid composition that in turn influence both structural and functional properties of cell membranes (Lyubenova & Schröder, 2010). It has been clearly demonstrated that lipoxygenase isoenzymes (LOX; EC 1.13.11.12) catalyse the dioxygenation of polyunsaturated fatty acids containing a cis,cis-1,4-pentadiene structure, with the consequent production of highly reactive and toxic hydroperoxy fatty acids (Figure 11.1). Subsequently, they are degraded into metabolites, such as jasmonates, conjugate dienoic acids and volatile aldehydes, such as malondialdehyde (MDA). LOX activity, nearly ubiquitous in the plant kingdom, is involved in stress responses, flowering, seed germination, pigment bleaching, flavour and aroma formation in plant products, and plant growth and development.



**Figure 11.1** The reaction catalysed by lipoxygenase indicates two possible products of linoleic acid degradation.

Despite numerous studies, LOX's physiological role in plants is not yet completely understood. In plants, there are mainly three isoenzymes: LOX-1, LOX-2 and LOX-3. The LOXs are globular proteins, soluble in water, constituted by a single polypeptide associated with an atom of Fe (III) that is essential for the catalysis. LOX isoenzymes, having different substrate specificity and optimum pH, are located in the cytosol, microsomes, plasma membrane and oil bodies. LOX-3 is the most abundant isoform and is the one that has the greatest activity (Sofo *et al.*, 2004a, and references within). For instance, the LOXs of olive trees are divided into soluble and membrane associated, the latter found mainly, but not exclusively, in the plastid fraction. Both in callus and in fruit of the olive tree, the main products of LOX are 13-hydroperoxides. The soluble isoforms are located in most of the tissues of olive trees, while the LOXs with acidic pH are associated with the chloroplast membranes of the fruit (Sofo *et al.*, 2004a, b).

Malondialdehyde has been utilized as a reliable biomarker of lipid peroxidation in plants (López *et al.*, 2011; Sofo *et al.*, 2004a, b; Sorkheh *et al.*, 2012a). Interestingly, MDA is also able to damage DNA, so enhancing the damage to cellular components and biomolecules.

The ROS produced in peroxisomes play a key role in plant ROS scavenging during abiotic stress conditions and senescence. Different experimental reports have indicated that peroxisomes are involved in cell signalling (by NO,  $O_2^{\bullet-}$ ,  $H_2O_2$  and S-nitrosoglutathione) during leaf senescence and in stress situations induced by xenobiotics and heavy metals. The activity of enzymes participating in the ascorbate-glutathione cycle in the peroxisomes is deeply influenced by abiotic stresses and senescence, when peroxisomal glutathione levels,  $H_2O_2$  concentration and lipid

peroxidation rate are considerably increased (Zentgraf, 2007). Furthermore, the peroxisomal NADH-dependent production of  $O_2^{\bullet-}$  radicals is enhanced by the reverse transition of leaf peroxisomes to glyoxysomes occurring when plants experience adverse environmental conditions (Hameed *et al.*, 2013; Sharma *et al.*, 2012).

### 11.3 Methodologies for lipid oxidation estimation

The hydroperoxide moiety of a LHP is reduced by divalent metal ions or glutathione-dependent peroxidases to an alcohol, with the production of a hydroxy derivative (LOH). In plants, many different products of lipid peroxidation can vary in the length of carbon chain and level of unsaturation of the hydroperoxy unoxidized PUFA composition. For this reason, simultaneous determination of both the substrate and its derivative oxidation products has been suggested (Browne & Armstrong, 2002).

Reverse-phase high-performance liquid chromatography (RP-HPLC) is an analytical technique capable of separating regioisomeric species of LHP and LOH derived from plant PUFA. Following total lipid extraction, alkaline hydrolysis and re-extraction of the liberated fatty acids, two separate systems with different mobile-phase conditions and analytical columns are usually used, one for LOH and LHP and the second for the native unoxidized PUFA. It was also reported (Browne & Armstrong, 2002) that a change of this methodology, allowing simultaneous determination of LHP, LOH and PUFA on a single chromatographic separation, using diode-array detection, allows determination of

the PUFA at 215 nm and the conjugated diene of LHP and LOH at 236 nm. This methodology sacrifices a small amount of resolution of LHP and LOH for inclusive determination of PUFAs in a single isocratic run but is useful for the determination of total LHP and LOH relative to their precursor PUFA within 20 min after injection.

Infrared spectroscopy (IS) can detect the major products of lipid peroxidation and is sensitive in detecting both hydroxyl and hydroperoxyl groups (Borchman & Sinha, 2002). This discrimination is especially useful for quantifying the oxidation of monounsaturated lipids, where secondary products of lipid oxidation are not promptly produced. Indeed, many plant lipids are highly unsaturated and so they are strongly subjected to lipid oxidation, particularly under stress conditions.

According to many authors (Sofa *et al.*, 2004a; Sorkheh *et al.* 2012a, b, and references within), MDA is usually extracted from plant tissues using trichloroacetic acid, and the supernatant resulting from centrifugation is added to thiobarbituric acid in 20% (w/v) trichloroacetic acid. The mixture obtained after heating (100°C) is subsequently cooled and centrifuged, and then absorbance values of the supernatant are recorded at 532, 600 and 440 nm. A series of compounds, including MDA, react with thiobarbituric acid, and for this they are called thiobarbituric acid-reactive substances (TBARS). When measuring MDA levels, especially in green tissues, it is very important to carry out a correction of the high sucrose content and also for the presence of anthocyanins or other interfering compounds. Thus, the values for a specific absorption at 600 nm are subtracted from the sample reading at 532 and 440 nm. Moreover, a standard curve of sucrose is used to correct the results from the interference of soluble sugars in samples.

## 11.4 Lipid oxidation in abiotic-stressed plants

Different types of abiotic stresses cause increase of ROS in plants, with consequent damage to lipids that ultimately results in oxidative stress (Foyer & Shigeoka, 2011; Gill & Tuteja, 2010; Miller *et al.*, 2010; Nishida and Murata, 1996; Sharma *et al.*, 2012). Plants under abiotic stresses must intercept photosynthetic light and at the same time avoid oxidative damage due to a specific

stressor or the combination of them. It is becoming increasingly clear that not only antioxidant enzymes and phenolic compounds but also soluble sugars (such as disaccharides, raffinose family, oligosaccharides and fructans), other compatible solutes (such as betaines and proline) and their associated metabolic enzymes are strong protective compounds against lipid peroxidation in stressed plants (Chen & Murata, 2002; Cruz *et al.*, 2013; Keunen *et al.*, 2013; Kotchoni *et al.*, 2006; Szabados & Saviouré, 2010).

### 11.4.1 Drought and salinity

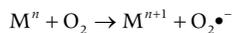
Membranes are considered to be a primary target of desiccation injury, as the ability of desiccation-tolerant organisms to avoid membrane damage during a dehydration-rehydration cycle is related to changes in membrane fluidity. Membrane bilayer structure in dried desiccation-tolerant organisms is thought to be stabilized as a result of interactions of the polar groups with sugars and proteins. Such interactions create space between phospholipids and prevent membrane phase changes. Membranes thus remain in the liquid-crystalline phase when the hydration shell is lost (Golovina & Hoekstra, 2003). Drought stress, especially if at a high degree and prolonged, is the main cause of photoinhibition, resulting in a light-dependent inactivation of the primary photochemistry associated with photosystem II. Though many species show a high tolerance to drought stress, net photosynthesis and transpiration rates generally decrease with increasing drought stress, whether for stomata closure or non-stomatal oxidative effects. Damage to lipid structure and functioning in plants subjected to various degrees of drought stress was recently found in cereals (Campo *et al.*, 2014; Csiszár *et al.*, 2012; Fukao *et al.*, 2011; Hameed *et al.*, 2011, 2013), various tree species (Štajner *et al.*, 2013), horticultural crop and forage plants (Abbas *et al.*, 2014; Slama *et al.*, 2011) and medicinal plants (Tian *et al.* 2012). Interestingly, exogenous cinnamic acid and derivatives of jasmonic acid were effectively used in improving plant drought stress tolerance by modulating the membrane lipid peroxidation and antioxidant activities (Anjum *et al.*, 2011; Sun *et al.*, 2012). In addition, Zhu *et al.* (2011) recently demonstrated that arbuscular mycorrhiza are able to alleviate the detrimental effect of drought by reducing MDA content and membrane permeability, and increasing proline content and antioxidant enzyme activities.

Drought, osmotic and salt stresses enhance the production of ROS, causing oxidative injury to lipids (Miller *et al.*, 2011, and references within). Salt stress, also at mild levels, is able to cause lipid peroxidation in cereals (Ashraf *et al.*, 2010; de Azevedo Neto *et al.*, 2006), herbs and vegetables (Sergio *et al.*, 2012; Tayebimeigooni *et al.*, 2012) and tree species (Ahmad *et al.*, 2010; Ayala-Astorga & Alcaraz-Meléndez, 2010). Tomato seedlings exposed to exogenous ascorbic acid show enhanced resistance against salt stress and decreased lipid peroxidation (Shalata & Neumann, 2001).

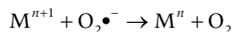
### 11.4.2 Metals

Elevated metal concentrations in the environment coincide with an oxidative stress-related constraint on platalidial and mitochondrial electron transport, which increases lipid peroxidation in these two compartments and in the whole cell (Dresler *et al.*, 2014; Keunen *et al.*, 2011; Nagajyoti *et al.*, 2010; Pospíšil, 2014; Yadav, 2010). Copper (Cu), iron (Fe), nickel (Ni), selenium (Se) and zinc (Zn), even if at relative low concentrations, are essential for plant physiological and biochemical processes. However, cadmium (Cd), aluminium (Al) and lead (Pb) are considered to be non-essential or toxic for plants (Cuyppers *et al.*, 2010; Sofo *et al.*, 2013).

Transition metals (M) are very good catalysts of oxygen reduction due to their unpaired electrons. Among these, Fe and Cu are the most important for plant metabolism.



In turn,  $O_2^{\bullet-}$  in aqueous solutions at neutral pH leads to the production of  $H_2O_2$ , subsequently decomposed to produce  $\bullet OH$  by the Haber–Weiss reaction, as follows:



Furthermore, transition metals lead to the formation of reactive alcohol radicals by the production of  $OH^-$  via the Fenton reaction:



Among metals, Cd alters the functionality of membranes by inducing lipid peroxidation and disturbs chloroplast metabolism by inhibiting chlorophyll biosynthesis and increasing photoinhibition (Ahmad *et al.*, 2011; Cuyppers *et al.*, 2010, 2011; Gallego *et al.*, 2012; Gill *et al.*, 2013; Pérez-Chaca *et al.*, 2014). On the other hand, Liptáková

*et al.* (2013) showed that the upregulation of LOXs is important for barley root response to toxic Cd but it is not responsible for the Cd-induced harmful lipid peroxidation. Also, Cu (Opdenakker *et al.*, 2012; Thounaojam *et al.*, 2012), Ni (Gajewska *et al.*, 2012; Kazemi *et al.*, 2010), Se (Malik *et al.*, 2012) and Pb (Maldonado-Magaña *et al.*, 2011), and other heavy metals and transition metals (Sytar *et al.*, 2013), are implicated in dose-specific and species-specific lipid peroxidation processes, and biological membranes are extremely susceptible to the presence of these metals into the soil.

## 11.5 Conclusion and future prospects

The status of membrane lipids can be perturbed by abiotic stressors, such as excess UV radiation, temperature extremes, nutrient deficiency and pollution of soil and air (Gill & Tuteja, 2010; Li *et al.*, 2012; Szarka *et al.*, 2012, and references within; Takshak & Agrawal, 2014; Tripathi *et al.*, 2011; Wyrwicka & Skłodowska, 2014; Yan *et al.*, 2010). The contribution of membrane lipids in protecting the photosynthetic machinery from photoinhibition during cold stress has been intensively discussed for many years (Nishida & Murata, 1996) and recently confirmed by Karabudak *et al.* (2014). In an important paper, Welti *et al.* (2002) conducted studies on membrane lipid profiles and on the role of some types of phospholipases in freezing-induced lipid change in *Arabidopsis*, demonstrating the crucial role of these enzymes in plant tolerance under cold conditions. Exposure of plants to herbicides appeared to be significantly related to the increase in lipid peroxidation (Dias *et al.*, 2014; McCarthy-Suárez *et al.*, 2011; Pazmiño *et al.*, 2011, Spoljaric *et al.*, 2011).

From the results of these researches, it appears clear that in the future the degree of lipid peroxidation in plants could be effectively used as a reliable biomarker of plant stresses, and of soil water and air pollution, with evident benefits for both agriculture and the environment.

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## References

- Abbas SR, Ahmad SD, Sabir SM, Shah AH (2014) Detection of drought tolerant sugarcane genotypes (*Saccharum officinarum*) using lipid peroxidation, antioxidant activity, glycine-betaine and proline contents. *J Soil Sci Plant Nutr* 14(1): doi: 10.4067/S0718-9516201400500001
- Ahmad P, Jaleel CA, Sharma S (2010) Antioxidant defense system, lipid peroxidation, proline-metabolizing enzymes, and biochemical activities in two *Morus alba* genotypes subjected to NaCl stress. *Russ J Plant Physiol* 57(4): 509–517.
- Ahmad P, Nabi G, Ashraf M (2011) Cadmium-induced oxidative damage in mustard [*Brassica juncea* (L.) Czern.& Coss.] plants can be alleviated by salicylic acid. *S Afr J Bot* 77: 36–44.
- Anjum SA, Wang L, Farooq M, Khan I, Xue L (2011) Methyl jasmonate-induced alteration in lipid peroxidation, antioxidative defence system and yield in soybean under drought. *J Agron Crop Sci* 97: 296–301.
- Ashraf MA, Ashraf A, Ali Q (2010) Response of two genetically diverse wheat cultivars to salt stress at different growth stages: leaf lipid peroxidation and phenolic content. *Pak J Bot* 42: 559–565.
- Ayala-Astorga GI, Alcaraz-Meléndez L (2010) Salinity effects on protein content, lipid peroxidation, pigments, and proline in *Paulownia imperialis* (Siebold & Zuccarini) and *Paulownia fortunei* (Seemann & Hemsley) grown *in vitro*. www.ejbiotechnology.info. doi: 10.2225/vol13-issue5-fulltext-13
- Borchman D, Sinha S (2002) Determination of products of lipid oxidation by infrared spectroscopy. In: Armstrong D (ed.), *Oxidative Stress Biomarkers and Antioxidant Protocols*. Humana Press, Totowa, New Jersey, pp. 21–28.
- Bradford KJ, Nonogaki H (2007) Lipid metabolism in seed dormancy. In: Penfield S, Pinfield-Wells H, Graham IA (eds), *Seed Development, Dormancy and Germination*. Blackwell Publishing, Oxford, pp. 133–152.
- Browne RW, Armstrong D (2002) Simultaneous determination of polyunsaturated fatty acids and corresponding monohydroperoxy and monohydroxy peroxidation products by HPLC. In: Armstrong D (ed.), *Oxidative Stress Biomarkers and Antioxidant Protocols*. Humana Press, Totowa, New Jersey, pp. 13–20.
- Campo S, Baldrich P, Meeguer J, Lalanne E, Coca M, San Segundo B (2014) Overexpression of a calcium-dependent protein kinase confers salt and drought tolerance in rice by preventing membrane lipid peroxidation. *Plant Physiol* 165: 688–704.
- Chen THH, Murata N (2002) Enhancement of tolerance of abiotic stress by metabolic engineering of betaines and other compatible solutes. *Curr Opin Plant Biol* 5: 250–257.
- Cruz FJR, Castro GLS, Silva Júnior DD, Festucci-Buselli RA, Pinheiro HA (2013) Exogenous glycine betaine modulates ascorbate peroxidase and catalase activities and prevent lipid peroxidation in mild water-stressed *Carapa guianensis* plants. *Photosynthetica* 51: 102–108.
- Csiszár J, Gallé Á, Horváth E, et al. (2012) Different peroxidase activities and expression of abiotic stress-related peroxidases in apical root segments of wheat genotypes with different drought stress tolerance under osmotic stress. *Plant Physiol Biochem* 52: 119–129.
- Cuttriss A, Pogson B (2004) Carotenoids. In: Dabies KM (ed.), *Plant Pigments and their Manipulation*. Blackwell Publishing, Oxford, pp. 57–91.
- Cuypers A, Plusquin M, Remans T, et al. (2010) Cadmium stress: an oxidative challenge. *Biometals* 23: 927–940.
- Cuypers A, Smeets K, Ruytinx J, et al. (2011) The cellular redox state as a modulator in cadmium and copper responses in *Arabidopsis thaliana* seedlings. *J Plant Physiol* 168: 309–316.
- De Azevedo Neto AD, Tarquinio Prisco J, Enéas-Filho J, Braga de Abreu CE, Gomes-Filho E (2006) Effect of salt stress on antioxidative enzymes and lipid peroxidation in leaves and roots of salt-tolerant and salt-sensitive maize genotypes. *Environ Exp Bot* 56: 87–94.
- Dias MC, Figueredo P, Duarte IF, Gil AM, Santos C (2014) Different responses of young and expanded lettuce leaves to fungicide Mancozeb: chlorophyll fluorescence, lipid peroxidation, pigments and proline content. *Photosynthetica* 52: 148–151.
- Dresler S, Hanaka A, Bednarek W, Maksymiec W (2014) Accumulation of low-molecular-weight organic acids in roots and leaf segments of *Zea mays* plants treated with cadmium and copper. *Acta Physiol Plant* 36: 1565–1575.
- Evert RF (2006) *Esau's Plant Anatomy. Meristems, Cells, and Tissues of the Plant Body: Their Structure, Function, and Development*, 3rd edn. John Wiley & Sons, New York.
- Foyer CH, Shigeoka S (2011) Understanding oxidative stress and antioxidant functions to enhance photosynthesis. *Plant Physiol* 155: 93–100.
- Fukao T, Yeung E, Bailey-Serres J (2011) The submergence tolerance regulator SUB1A mediates crosstalk between submergence and drought tolerance in rice. *Plant Cell* 23: 412–427.
- Gajewska E, Bernat P, Długoński J, Skłodowski M (2012) Effect of nickel on membrane integrity, lipid peroxidation and fatty acid composition in wheat seedlings. *J Agron Crop Sci* 198: 286–294.
- Gallego SM, Pena LB, Barcia RA, et al. (2012) Unravelling cadmium toxicity and tolerance in plants: insight into regulatory mechanisms. *Environ Exp Bot* 83: 33–46.
- Gill SS, Hasanuzzaman M, Nahar K, Macovei A, Tuteja N (2013) Importance of nitric oxide in cadmium stress tolerance in crop plants. *Plant Physiol Biochem* 63: 254–261.
- Golovina EA, Hoekstra FA (2003) Structural changes in membranes of developing wheat embryos during the acquisition of desiccation tolerance. In: Nicolás G, Bradford KJ, Côme D, Pritchard HW (eds), *The Biology of Seeds*. Recent Research Advances. CAB International, Wallingford, pp. 337–344.
- Hameed A, Bibi N, Akhter J, Iqbal N (2011) Differential changes in antioxidants, proteases, and lipid peroxidation in flag leaves of wheat genotypes under different levels of water deficit conditions. *Plant Physiol Biochem* 49: 178–185.
- Hameed A, Goher M, Iqbal N (2013) Drought induced programmed cell death and associated changes in antioxidants,

- proteases, and lipid peroxidation in wheat leaves. *Biol Plant* 57: 370–374.
- Hinojosa MB, García-Ruiz R, Carreira JA (2010) Utilizing microbial community structure and function to evaluate the health of heavy metal polluted soils. In: Sherameti I, Varna A (eds), *Soil Heavy Metals*. Soil Biology, vol. 19. Springer-Verlag, Berlin pp. 185–224.
- Karabudak T, Bor M, Özdemir F, Türkan İ (2014) Glycine betaine protects tomato (*Solanum lycopersicum*) plants at low temperature by inducing fatty acid desaturase7 and lipoxygenase gene expression. *Mol Biol Rep* 42: 1401–1410.
- Kazemi N, Khavari-Nejad RA, Fahimi H, Saadatmand S, Nejad-Sattari T (2010) Effects of exogenous salicylic acid and nitric oxide on lipid peroxidation and antioxidant enzyme activities in leaves of *Brassica napus* L. under nickel stress. *Sci Hort* 126: 402–407.
- Keunen E, Remans T, Bohler S, Vangronsveld J, Cuypers A (2011) Metal-induced oxidative stress and plant mitochondria. *Int J Mol Sci* 12: 6894–6918.
- Keunen E, Peshev D, Vangronsveld J, van den Ende W, Cuypers A (2013) Plant sugars are crucial players in the oxidative challenge during abiotic stress: extending the traditional concept. *Plant Cell Environ* 36: 1242–1255.
- Kotchoni SO, Kuhns C, Ditzer A, Kirch H-H, Bartels D (2006) Over-expression of different aldehyde dehydrogenase genes in *Arabidopsis thaliana* confers tolerance to abiotic stress and protects plants against lipid peroxidation and oxidative stress. *Plant Cell Environ* 29: 1033–1048.
- Li X, Zhang L, Li Y, Ma L, Bu N, Ma C (2012) Changes in photosynthesis, antioxidant enzymes and lipid peroxidation in soybean seedlings exposed to UV-B radiation and/or Cd. *Plant Soil* 352: 377–387.
- Liptáková Ľ, Huttová J, Mistrík I, Tamás L (2013) Enhanced lipoxygenase activity is involved in the stress response but not in the harmful lipid peroxidation and cell death of short-term cadmium-treated barley root tip. *J Plant Physiol* 170: 646–652.
- López MA, Vicente J, Kulasekaran S, et al. (2011) Antagonistic role of 9-lipoxygenase-derived oxylipins and ethylene in the control of oxidative stress, lipid peroxidation and plant defence. *Plant J* 67: 447–458.
- Lyubenova L, Schröder P (2010) Uptake and effect of heavy metals on the plant detoxification cascade in the presence and absence of organic pollutants. In: Sherameti I, Varna A (eds), *Soil Heavy Metals*. Soil Biology, vol. 19. Springer-Verlag, Berlin, pp. 65–85.
- Maldonado-Magaña A, Favela-Torres E, Rivera-Cabrera F, Volke-Sepulveda TL (2011) Lead bioaccumulation in *Acacia farnesiana* and its effect on lipid peroxidation and glutathione production. *Plant Soil* 339: 377–389.
- Malik JA, Goel S, Kaur N, Sharma S, Singh I, Nayyara H (2012) Selenium antagonises the toxic effects of arsenic on mungbean (*Phaseolus aureus* Roxb.) plants by restricting its uptake and enhancing the antioxidative and detoxification mechanisms. *Environ Exp Bot* 77: 242–248.
- McCarthy-Suárez I, Gómez M, del Río LA, Palma JM (2011) Organ-specific effects of the auxin herbicide 2,4-D on the oxidative stress and senescence-related parameters of the stems of pea plants. *Acta Physiol Plant* 33: 2239–2247.
- Miller G, Suzuki N, Ciftci-Yilmaz S, Mittler R (2010) Reactive oxygen species homeostasis and signaling during drought and salinity stresses. *Plant Cell Environ* 33: 453–467.
- Nagajyoti PC, Lee KD, Srekanth TVM (2010) Heavy metals, occurrence and toxicity for plants: a review. *Environ Chem Lett* 8: 199–216.
- Nishida I, Murata N (1996) Chilling sensitivity in plants and cyanobacteria: the crucial contribution of membrane lipids. *Annu Rev Plant Physiol Plant Mol Biol* 47: 541–568.
- Opendakker K, Remans T, Keunen E, Vangronsveld J, Cuypers A (2012) Exposure of *Arabidopsis thaliana* to Cd or Cu excess leads to oxidative stress mediated alterations in MAPKinase transcript levels. *Environ Exp Bot* 83: 53–61.
- Pazmiño DM, Rodríguez-Serrano M, Romero-Puertas MC, Archilla-Ruiz A, del Río LA, Sandalio LM (2011) Differential response of young and adult leaves to herbicide 2,4-dichlorophenoxyacetic acid in pea plants: role of reactive oxygen species. *Plant Cell Environ* 34: 1874–1889.
- Pérez-Chaca MV, Rodríguez-Serrano M, Molina AS, et al. (2014) Cadmium induces two waves of reactive oxygen species in *Glycine max* (L.) roots. *Plant Cell Environ* 37: 1672–1687.
- Pospišil P (2014) The role of metals in production and scavenging of reactive oxygen species in Photosystem II. *Plant Cell Physiol* 55: 1224–1232.
- Robinson C, Mant A (2005) Biogenesis of the thylakoid membrane. In: Møller SG (ed.), *Plastids*. Annual Plant Reviews, vol. 13. Blackwell Publishing, Oxford, pp. 180–213.
- Selmar D (2010) Biosynthesis of cyanogenic glycosides, glucosinolates and non-protein amino acid. In: Wink M (ed.), *Biochemistry of Plant Secondary Metabolism*, 2nd edn. Annual Plant Reviews, vol. 40. Wiley-Blackwell, Oxford, pp. 92–181.
- Sergio L, de Paola A, Cantore V, et al. (2012) Effect of salt stress on growth parameters, enzymatic antioxidant system, and lipid peroxidation in wild chicory (*Cichorium intybus* L.). *Acta Physiol Plant* 34: 2349–2358.
- Shalata A, Neumann PM (2001) Exogenous ascorbic acid (vitamin C) increases resistance to salt stress and reduces lipid peroxidation. *J Exp Bot* 52: 2207–2211.
- Sharma P, Bhushan Jha A, Shanker Dubey R, Pessarikli M (2012) Reactive oxygen species, oxidative damage, and antioxidative defense mechanism in plants under stressful conditions. *J Bot* 2012: article ID 217037. doi:10.1155/2012/217037
- Singh Gill S, Tuteja N (2010) Reactive oxygen species and antioxidant machinery in abiotic stress tolerance in crop plants. *Plant Physiol Biochem* 48: 909–930.
- Slama I, Tayachi S, Jdey A, Rouached A, Abdelly C (2011) Differential response to water deficit stress in alfalfa (*Medicago sativa*) cultivars: growth, water relations, osmolyte accumulation and lipid peroxidation. *Afr J Biotechnol* 10: 16250–16259.

- Sofo A, Dichio B, Xiloyannis C, Masia A (2004a) Lipoxygenase activity and proline accumulation in leaves and roots of olive tree in response to drought stress. *Physiol Plant* 121: 58–65.
- Sofo A, Dichio B, Xiloyannis C, Masia A (2004b) Effects of different irradiance levels on some antioxidant enzymes and on malondialdehyde content during rewatering in olive tree. *Plant Sci* 166: 293–302.
- Sofo A, Vitti A, Nuzzaci M, et al. (2013) Correlation between hormonal homeostasis and morphogenic responses in *Arabidopsis thaliana* seedlings growing in a Cd/Cu/Zn multi-pollution context. *Physiol Plant* 149: 487–498.
- Sorkheh K, Shiran B, Rouhi V, Khodambashi M, Sofo A (2012a) Salt stress induction of some key antioxidant enzymes and metabolites in eight Iranian wild almond species. *Acta Physiol Plant*: 203–213.
- Sorkheh K, Shiran B, Khodambashi M, Rouhi V, Mosavei S, Sofo A (2012b) Exogenous proline alleviates the effects of H<sub>2</sub>O<sub>2</sub>-induced oxidative stress in wild almond species (*Prunus* spp.). *Russ J Plant Physiol* 59: 788–798.
- Spoljaric D, Cipak A, Horvatic J, et al. (2011) Endogenous 4-hydroxy-2-nonenal in microalga *Chlorella kessleri* acts as a bioactive indicator of pollution with common herbicides and growth regulating factor of hormesis. *Aquat Toxic* 105: 552–558.
- Štajner D, Orlović S, Popović BM, Kebert M, Galić Z (2011) Screening of drought oxidative stress tolerance in Serbian melliferous plant species. *Afr J Biotechnol* 10: 1609–1614.
- Sun WJ, Nie YX, Gao Y, Dai A, Bai JG (2012) Exogenous cinnamic acid regulates antioxidant enzyme activity and reduces lipid peroxidation in drought-stressed cucumber leaves. *Acta Physiol Plant* 34: 641–655.
- Sytar O, Kumar A, Latowski D, Kuczynska P, Strzałka K, Prasad MNV (2013) Heavy metal-induced oxidative damage, defense reactions, and detoxification mechanisms in plants. *Acta Physiol Plant* 35: 985–999.
- Szabados L, Savouré A (2010) Proline: a multifunctional amino acid. *Trends Plant Sci* 15: 89–97.
- Szarka A, Tomasskovic B, Bánhegyi G (2012) The ascorbate-glutathione- $\alpha$ -tocopherol triad in abiotic stress response. *Int J Mol Sci* 13: 4458–4483.
- Takshak S, Agrawal SB (2014) Effect of ultraviolet-B radiation on biomass production, lipid peroxidation, reactive oxygen species, and antioxidants in *Withania somnifera*. *Biol Plant* 58: 328–334.
- Tayebimeigooni A, Awang Y, Mahmood M, Selamat A, Wahab Z (2012) Leaf water status, proline content, lipid peroxidation and accumulation of hydrogen peroxide in salinized Chinese kale (*Brassica alboglabra*). *J Food Agric Environ* 10: 371–374.
- Tetlow IJ, Rawsthorne S, Raines C, Emes MJ (2005) Plastid metabolic pathways. In: Møller SG (ed.) *Plastids*. Annual Plant Reviews, vol. 13. Blackwell Publishing, Oxford, pp. 60–125.
- Thounaojam TC, Panda P, Mazumdar P, et al. (2012) Excess copper induced oxidative stress and response of antioxidants in rice. *Plant Physiol Biochem* 53: 33–39.
- Tian Z, Wang F, Zhang W, Liu C, Zhao X (2012) Antioxidant mechanism and lipid peroxidation patterns in leaves and petals of marigold in response to drought Stress. *Hort Environ Biotechnol* 53: 183–192.
- Tripathi R, Sarkar A, Rai SP, Agrawal SB (2011) Supplemental ultraviolet-B and ozone: impact on antioxidants, proteome and genome of linseed (*Linum usitatissimum* L. cv. Padmini). *Plant Biol* 13: 93–104.
- Welti R, Li W, Li M, et al. (2002) Profiling membrane lipids in plant stress responses: role of phospholipase D $\alpha$  in freezing-induced lipid change in *Arabidopsis*. *J Biol Chem* 277: 31994–30002.
- Wyrwicka A, Skłodowska M (2014) Intercompartmental differences between cytosol and mitochondria in their respective antioxidative responses and lipid peroxidation levels in acid rain stress. *Acta Physiol Plant* 36: 837–848.
- Yadav K (2010) Heavy metals toxicity in plants: an overview on the role of glutathione and phytochelatin in heavy metal stress tolerance of plants. *S Afr J Bot* 76: 167–179.
- Yan K, Chen W, He X, Zhang G, Xu S Wang L (2010) Responses of photosynthesis, lipid peroxidation and antioxidant system in leaves of *Quercus mongolica* to elevated O<sub>3</sub>. *Environ Exp Bot* 69: 198–204.
- Zentgraf U (2007) Oxidative stress and leaf senescence. In: Gan S (ed.), *Senescence Processes in Plants*. Blackwell Publishing, Oxford, pp. 69–86.
- Zhu X, Song F, Liu S (2011) Arbuscular mycorrhiza impacts on drought stress of maize plants by lipid peroxidation, proline content and activity of antioxidant system. *J Food Agric Environ* 9: 583–587.